

CRUSTACEAN ISSUES 18



Decapod Crustacean Phylogenetics

edited by

Joel W. Martin, Keith A. Crandall, and Darryl L. Felder



CRC Press
Taylor & Francis Group

Decapod Crustacean Phylogenetics

Edited by

Joel W. Martin

Natural History Museum of L. A. County
Los Angeles, California, U. S. A.

Keith A. Crandall

Brigham Young University
Provo, Utah, U. S. A.

Darryl L. Felder

University of Louisiana
Lafayette, Louisiana, U. S. A.



CRC Press

Taylor & Francis Group

Boca Raton London New York

CRC Press is an imprint of the
Taylor & Francis Group, an **informa** business

CRC Press
Taylor & Francis Group
6000 Broken Sound Parkway NW, Suite 300
Boca Raton, FL 33487-2742

© 2009 by Taylor & Francis Group, LLC
CRC Press is an imprint of Taylor & Francis Group, an Informa business

No claim to original U.S. Government works
Printed in the United States of America on acid-free paper
10 9 8 7 6 5 4 3 2 1

International Standard Book Number-13: 978-1-4200-9258-5 (Hardcover)

This book contains information obtained from authentic and highly regarded sources. Reasonable efforts have been made to publish reliable data and information, but the author and publisher cannot assume responsibility for the validity of all materials or the consequences of their use. The authors and publishers have attempted to trace the copyright holders of all material reproduced in this publication and apologize to copyright holders if permission to publish in this form has not been obtained. If any copyright material has not been acknowledged please write and let us know so we may rectify in any future reprint.

Except as permitted under U.S. Copyright Law, no part of this book may be reprinted, reproduced, transmitted, or utilized in any form by any electronic, mechanical, or other means, now known or hereafter invented, including photocopying, microfilming, and recording, or in any information storage or retrieval system, without written permission from the publishers.

For permission to photocopy or use material electronically from this work, please access www.copyright.com (<http://www.copyright.com/>) or contact the Copyright Clearance Center, Inc. (CCC), 222 Rosewood Drive, Danvers, MA 01923, 978-750-8400. CCC is a not-for-profit organization that provides licenses and registration for a variety of users. For organizations that have been granted a photocopy license by the CCC, a separate system of payment has been arranged.

Trademark Notice: Product or corporate names may be trademarks or registered trademarks, and are used only for identification and explanation without intent to infringe.

Library of Congress Cataloging-in-Publication Data

Decapod crustacean phylogenetics / editors, Joel W. Martin, Keith A. Crandall, Darryl L. Felder.
p. cm. -- (Crustacean issues)

Includes bibliographical references and index.

ISBN 978-1-4200-9258-5 (hardcover : alk. paper)

1. Decapoda (Crustacea) 2. Phylogeny. I. Martin, Joel W. II. Crandall, Keith A. III. Felder, Darryl L.
IV. Title. V. Series.

Q1.444.M33D44 2009

595.3'8138--dc22

2009001091

Visit the Taylor & Francis Web site at
<http://www.taylorandfrancis.com>

and the CRC Press Web site at
<http://www.crcpress.com>

Contents

Preface	ix
JOEL W. MARTIN, KEITH A. CRANDALL & DARRYL L. FELDER	
I <i>Overviews of Decapod Phylogeny</i>	
On the Origin of Decapoda	3
FREDERICK R. SCHRAM	
Decapod Phylogenetics and Molecular Evolution	15
ALICIA TOON, MAEGAN FINLEY, JEFFREY STAPLES & KEITH A. CRANDALL	
Development, Genes, and Decapod Evolution	31
GERHARD SCHOLTZ, ARKHAT ABZHANOV, FREDERIKE ALWES, CATERINA BIFFIS & JULIA PINT	
Mitochondrial DNA and Decapod Phylogenies: The Importance of Pseudogenes and Primer Optimization	47
CHRISTOPH D. SCHUBART	
Phylogenetic Inference Using Molecular Data	67
FERRAN PALERO & KEITH A. CRANDALL	
Decapod Phylogeny: What Can Protein-Coding Genes Tell Us?	89
K.H. CHU, L.M. TSANG, K.Y. MA, T.Y. CHAN & P.K.L. NG	
Spermatozoal Morphology and Its Bearing on Decapod Phylogeny	101
CHRISTOPHER TUDGE	
The Evolution of Mating Systems in Decapod Crustaceans	121
AKIRA ASAKURA	
A Shrimp's Eye View of Evolution: How Useful Are Visual Characters in Decapod Phylogenetics?	183
MEGAN L. PORTER & THOMAS W. CRONIN	
Crustacean Parasites as Phylogenetic Indicators in Decapod Evolution	197
CHRISTOPHER B. BOYKO & JASON D. WILLIAMS	
The Bearing of Larval Morphology on Brachyuran Phylogeny	221
PAUL F. CLARK	

II *Advances in Our Knowledge of Shrimp-Like Decapods*

- Evolution and Radiation of Shrimp-Like Decapods: An Overview 245
CHARLES H.J.M. FRANSEN & SAMMY DE GRAVE

- A Preliminary Phylogenetic Analysis of the Dendrobranchiata Based on Morphological Characters 261
CAROLINA TAVARES, CRISTIANA SEREJO & JOEL W. MARTIN

- Phylogeny of the Infraorder Caridea Based on Mitochondrial and Nuclear Genes (Crustacea: Decapoda) 281
HEATHER D. BRACKEN, SAMMY DE GRAVE & DARRYL L. FELDER

III *Advances in Our Knowledge of the Thalassinidean and Lobster-Like Groups*

- Molecular Phylogeny of the Thalassinidea Based on Nuclear and Mitochondrial Genes 309
RAFAEL ROBLES, CHRISTOPHER C. TUDGE, PETER C. DWORSCHAK, GARY C.B. POORE & DARRYL L. FELDER

- Molecular Phylogeny of the Family Callinassidae Based on Preliminary Analyses of Two Mitochondrial Genes 327
DARRYL L. FELDER & RAFAEL ROBLES

- The Timing of the Diversification of the Freshwater Crayfishes 343
JESSE BREINHOLT, MARCOS PÉREZ-LOSADA & KEITH A. CRANDALL

- Phylogeny of Marine Clawed Lobster Families Nephropidae Dana, 1852, and Thaumastocheilidae Bate, 1888, Based on Mitochondrial Genes 357
DALE TSHUDY, RAFAEL ROBLES, TIN-YAM CHAN, KA CHAI HO, KA HOU CHU, SHANE T. AHYONG & DARRYL L. FELDER

- The Polychelidan Lobsters: Phylogeny and Systematics (Polychelida: Polychelidae) 369
SHANE T. AHYONG

IV *Advances in Our Knowledge of the Anomura*

- Anomuran Phylogeny: New Insights from Molecular Data 399
SHANE T. AHYONG, KAREEN E. SCHNABEL & ELIZABETH W. MAAS

V *Advances in Our Knowledge of the Brachyura*

- Is the Brachyura Podotremata a Monophyletic Group? 417
GERHARD SCHOLTZ & COLIN L. MCLAY

Assessing the Contribution of Molecular and Larval Morphological Characters in a Combined Phylogenetic Analysis of the Superfamily Majoidea	437
KRISTIN M. HULTGREN, GUILLERMO GUERAO, FERNANDO P.L. MARQUES & FERRAN P. PALERO	
Molecular Genetic Re-Examination of Subfamilies and Polyphyly in the Family Pinnotheridae (Crustacea: Decapoda)	457
EMMA PALACIOS-THEIL, JOSÉ A. CUESTA, ERNESTO CAMPOS & DARRYL L. FELDER	
Evolutionary Origin of the Gall Crabs (Family Cryptochiridae) Based on 16S rDNA Sequence Data	475
REGINA WETZER, JOEL W. MARTIN & SARAH L. BOYCE	
Systematics, Evolution, and Biogeography of Freshwater Crabs	491
NEIL CUMBERLIDGE & PETER K.L. NG	
Phylogeny and Biogeography of Asian Freshwater Crabs of the Family Gecarcinucidae (Brachyura: Potamoidea)	509
SEBASTIAN KLAUS, DIRK BRANDIS, PETER K.L. NG, DARREN C.J. YEO & CHRISTOPH D. SCHUBART	
A Proposal for a New Classification of Portunoidea and Cancroidea (Brachyura: Heterotremata) Based on Two Independent Molecular Phylogenies	533
CHRISTOPH D. SCHUBART & SILKE REUSCHEL	
Molecular Phylogeny of Western Atlantic Representatives of the Genus <i>Hexapanopeus</i> (Decapoda: Brachyura: Panopeidae)	551
BRENT P. THOMA, CHRISTOPH D. SCHUBART & DARRYL L. FELDER	
Molecular Phylogeny of the Genus <i>Cronius</i> Stimpson, 1860, with Reassignment of <i>C. tumidulus</i> and Several American Species of <i>Portunus</i> to the Genus <i>Achelous</i> De Haan, 1833 (Brachyura: Portunidae)	567
FERNANDO L. MANTELATTO, RAFAEL ROBLES, CHRISTOPH D. SCHUBART & DARRYL L. FELDER	
Index	581
Color Insert	

Evolutionary Origin of the Gall Crabs (Family Cryptochiridae) Based on 16S rDNA Sequence Data

REGINA WETZER¹, JOEL W. MARTIN¹ & SARAH L. BOYCE²

¹ Natural History Museum of Los Angeles County, 900 Exposition Boulevard, Los Angeles, CA 90007

² Harvard University, Cambridge, MA 02138

ABSTRACT

Gall crabs (family Cryptochiridae) are small brachyuran crabs living on or in depressions formed in scleractinian corals. Their adaptation to this unusual habitat has led to specializations, including mucous feeding, small body size, and relatively short appendages. Currently, gall crabs are treated as constituting a distinct superfamily (Cryptochiroidea) that contains the sole family Cryptochiridae. There has never been an attempt to elucidate the relationships of the gall crabs to other brachyurans. The group is therefore an ideal candidate for employing molecular data to deduce phylogenetic relationships. We sequenced a 545-bp fragment of the 16S mitochondrial gene from specimens of a widespread species of cryptochirid (*Hapalocarcinus marsupialis*) from Mexico and French Polynesia and compared these to other crab sequences available in GenBank. Our preliminary analyses confirm the placement of the cryptochirids in the Brachyura subsection Thoracotremata. Our results also indicate that cryptochirids are members of the superfamily Grapsoidea and are probably closely allied with the family Grapsidae. The Grapsoidea as presently defined is considered a paraphyletic assemblage.

1 INTRODUCTION

Crabs of the family Cryptochiridae Paul'son, 1875, are among the most unusual of all groups of decapod crustaceans. From what little we know about their biology and natural history, it appears that young crabs settle on scleractinian corals, and most species somehow induce the coral to grow over and around the crab. For some cryptochirids, the result is merely a protective indentation or crevice within the coral, and there appears to be little modification of the host. Females, and in some cases males, live in open pits or tunnels in the corals, or on the surface of the corals. Some species (notably *Hapalocarcinus marsupialis* and *Pseudohapalocarcinus ransoni*) live within the protective confines of a coral "gall" that completely or partially (in the case of *Pseudohapalocarcinus*) encompasses and protects the crab, where it remains for the remainder of its life (see Kropp 1986, 1988; Abelson et al. 1991; Carricart-Ganivet et al. 2004 for reviews of species-specific life histories). Males, which are far smaller than females, and about which less is generally known, are also sometimes found in pits or depressions on the same coral (e.g., the crab genus *Fungicola*, which inhabits fungiid corals) or are not directly associated with the coral as far as is known. Currently, the family includes 46 extant species (there are no known fossil species) partitioned among 20 genera (Table 1; see also Ng et al. 2008: 212). Cryptochirids are probably found wherever scleractinian coral reefs occur worldwide, although some reef systems have yet to be rigorously sampled for them. There are also species associated with deep-water, ahermatypic corals found far from reefs. Although roughly circumtropical in distribution, the group is most diverse in the Indo-West Pacific. Table 1 is the first compilation

Table 1. Comprehensive list of described genera (in bold) and species of the family Cryptochiridae, with a summary of the coral families and genera that the crabs inhabit, general biogeographic distributions of the crab genera, and depth records. Depth applies to the entire geographic range.

Genus and Species	Known Coral Hosts	General Distribution (of crab)	Primary References
<i>Cecidocarcinus</i> Kropp & Manning, 1987			
	Dendrophyllidae: <i>Dendrophyllia</i> , <i>Enallopsammia</i>	Atlantic: Valdivia Ridge (southeastern Atlantic, off Namibia); depth 512 m	Kropp & Manning 1987
<i>Cecidocarcinus brychius</i> Kropp & Manning, 1987			
<i>Cecidocarcinus zibrowii</i> Manning, 1991			
<i>Cryptochirus</i> Heller, 1861			
	Faviidae: <i>Cyphastrea</i> , <i>Barabatoia</i> , <i>Favia</i> , <i>Favites</i> , <i>Goniastrea</i> , <i>Leptoria</i> , <i>Montastrea</i> , <i>Platygyra</i>	Red Sea Pacific: Vietnam, Japan, Micronesia (Palau, Guam, Pohnpei); depth <1 to 30 m	Kropp 1990 Wei et al. 2006
	Oculinidae: <i>Cyathelia</i>		
<i>Cryptochirus coralliodytes</i> Heller, 1861			
<i>Cryptochirus planus</i> (Takeda & Tamura, 1983)			
<i>Cryptochirus rubrilineatus</i> Fize & Serène, 1957			
<i>Dacryomaia</i> Kropp, 1990			
	Siderastreidae: <i>Psammocora</i>	Pacific: Vietnam, Japan (Isu Islands, Ogasawara Islands, Ryukyu Islands), Micronesia (Palau, Guam); depth <1 to 8 m	Kropp 1990 Wei et al. 2006
<i>Dacryomaia edmondsoni</i> (Fize & Serène, 1956a)			
<i>Dacryomaia japonica</i> (Takeda & Tamura, 1981b)			
<i>Dacryomaia</i> sp. 1		Pacific: Micronesia (Guam)	Paulay et al. 2003
<i>Dacryomaia</i> sp. 2		Pacific: Micronesia (Guam)	Paulay et al. 2003
<i>Detocarcinus</i> Kropp & Manning, 1987			
	Caryophyllidae: <i>Asterosimilia</i> , <i>Caryophyllia</i>	Atlantic: off Ghana	Kropp & Manning 1987
	Dendrophyllidae: <i>Dendrophyllia</i> (questionable)		
	Oculinidae: <i>Schizoculina</i>		
	Rhizangiidae: <i>Phyllangia</i>		
<i>Detocarcinus balssi</i> (Monod, 1956)			

Table 1. continued.

Genus and Species	Known Coral Hosts	General Distribution (of crab)	Primary References
<i>Fizesereneia</i> Takeda & Tamura, 1980b			
Mussidae: <i>Acanthastrea</i> , <i>Lobophyllia</i> , <i>Symphyllia</i>		Pacific: Vietnam, Indonesia, Japan (Izu Islands, Ryukyu Islands), Australia, Micronesia (Palau, Guam, Pohnpei); depth 1 to 15 m	Kropp 1990
<i>Fizesereneia heimi</i> (Fize & Serène, 1956a)			
<i>Fizesereneia ishikawai</i> (Takeda & Tamura, 1980b)			
<i>Fizesereneia latisella</i> Kropp, 1994			
<i>Fizesereneia stimpsoni</i> (Fize & Serène, 1956b)			
<i>Fizesereneia tholia</i> Kropp, 1994			
<i>Fungicola</i> Serène, 1966			
Fungiidae: <i>Fungia</i> , <i>Podobacia</i> , <i>Sandalolitha</i>		Pacific: Vietnam, Indonesia, Japan (Ryukyu Islands), Micronesia (Palau, Guam); depth 1 to 15 m	Kropp 1990
<i>Fungicola fagei</i> (Fize & Serène, 1956a)			
<i>Fungicola utinomii</i> (Fize & Serène, 1956a)			
<i>Hapalocarcinus</i> Stimpson, 1859			
Pocilloporidae: <i>Pocillopora</i> , <i>Seriatopora</i> , <i>Stylophora</i>		Pacific: Indo-West Pacific to Eastern Pacific (Colombia) Red Sea; depth 1 to 27 m	Kropp 1990 Wei et al. 2006
<i>Hapalocarcinus marsupialis</i> Stimpson, 1859			
<i>Hiroia</i> Takeda & Tamura, 1981a			
Faviidae: <i>Cyphastrea</i> , <i>Hydnophora</i>		Pacific: Vietnam, Japan (Izu Islands, Ryukyu Islands), Micronesia (Palau, Guam); depth 1 to 19 m	Kropp 1990 Wei et al. 2006
Merulinidae: <i>Merulina</i>			

Table 1. continued.

Genus and Species	Known Coral Hosts	General Distribution (of crab)	Primary References
<i>Hiroia krempfi</i> (Fize & Serène, 1956a)	<i>Lithoscaptus</i> Milne Edwards, 1862 Faviidae: <i>Cyphastrea</i> , <i>Echinopora</i> , <i>Favia</i> , <i>Favites</i> , <i>Hydnophora</i> , <i>Goniastrea</i> , <i>Leptastrea</i> , <i>Platygyra</i> , <i>Plesiastrea</i> Merulinidae: <i>Merulina</i>	Pacific: Réunion, Vietnam, Japan (Izu Islands, Kushimoto, Ogasawara Islands, Ryukyu Islands), Micronesia (Palau, Guam, Pohnpei), Palmyra Island, Teraina; depth <1 to 12 m	Kropp 1990 Wei et al. 2006
<i>Lithoscaptus grandis</i> (Takeda & Tamura, 1983)	<i>Lithoscaptus helleri</i> (Fize & Serène, 1957)		
<i>Lithoscaptus nami</i> (Fize & Serène, 1957)			
<i>Lithoscaptus</i> (?) <i>pacificus</i> (Edmondson, 1933) ¹			
<i>Lithoscaptus paradoxus</i> Milne Edwards, 1862			
<i>Lithoscaptus pardalotus</i> Kropp, 1995			
<i>Lithoscaptus prionotus</i> Kropp, 1994			
<i>Lithoscaptus tri</i> (Fize & Serène, 1956b)			
<i>Luciades</i> Kropp & Manning, 1996	Pavonidae: <i>Leptoseria</i>	Pacific: Micronesia (Guam); depth 128 to 137 m	Kropp & Manning 1996
<i>Luciades agana</i> Kropp & Manning, 1996			
<i>Neotroglocarcinus</i> Takeda & Tamura, 1980a	Dendrophyllidae: <i>Turbinaria</i>	Pacific: Vietnam, Japan (Izu Islands, Ryukyu Islands), Micronesia (Palau, Guam, Pohnpei), Enewetak, Hong Kong; depth <1 to 13 m	Kropp 1990 Wei et al. 2006
<i>Neotroglocarcinus hongkongensis</i> (Shen, 1936)			
<i>Neotroglocarcinus dawydoffi</i> (Fize & Serène, 1956a)			

Table 1. continued.

Genus and Species	Known Coral Hosts	General Distribution (of crab)	Primary References
<i>Opecarcinus</i> Kropp & Manning, 1987	Agariciidae: <i>Agaricia</i> , <i>Gardineroseris</i> , <i>Leptoseris</i> , <i>Pavona</i> Siderasteriidae: <i>Coscinaraea</i> , <i>Siderastrea</i>	Pacific: Vietnam, Japan, to west coast of Mexico Indian Ocean: Christmas Island Atlantic Ocean: Ascension Island and western Atlantic (Caribbean, Gulf of Mexico south to Brazil); depth <1 to 82 m	Kropp & Manning 1987 Kropp 1990 Wei et al. 2006
<i>Opecarcinus aurantius</i> Kropp, 1989			
<i>Opecarcinus crescentus</i> (Edmondson, 1925)			
<i>Opecarcinus granulatus</i> (Shen, 1936)			
<i>Opecarcinus hypostegus</i> (Shaw & Hopkins, 1977)			
<i>Opecarcinus lobifrons</i> Kropp, 1989			
<i>Opecarcinus peliops</i> Kropp, 1989			
<i>Opecarcinus pholeter</i> Kropp, 1989			
<i>Opecarcinus sierra</i> Kropp, 1989			
<i>Pelycomaia</i> Kropp, 1990	Faviidae: <i>Cyphastrea</i> , <i>Leptastrea</i>	Pacific: Vietnam, Micronesia (Guam), Hawaii; depth < 2 m	Kropp 1990
<i>Pelycomaia minuta</i> (Edmondson, 1933)			
<i>Pseudocryptochirus</i> Hiro, 1938	Dendrophyllidae: <i>Turbinaria</i>	Pacific: Vietnam, Indonesia, Japan (Isu Islands), Micronesia (Palau, Guam, Pohnpei); depth 1 to 6 m	Kropp 1990 Wei et al. 2006
<i>Pseudocryptochirus viridis</i> Hiro, 1938			

Table 1. continued.

Genus and Species	Known Coral Hosts	General Distribution (of crab)	Primary References
<i>Pseudohapalocarcinus</i> Fize & Serène, 1956a	Agariciidae: <i>Pavona</i>	Pacific: Vietnam, Japan (Ryukyu Islands), Micronesia (Palau, Guam, Pohnpei); depth <1 to 21 m	Kropp 1990
<i>Pseudohapalocarcinus ransoni</i> Fize & Serène, 1956a			
<i>Sphenomaia</i> Kropp, 1990	Faviidae: <i>Favites</i> , <i>Hydnophora</i> , <i>Platygyra</i>	Central Pacific (Teraina); depth not recorded	Kropp 1990
<i>Sphenomaia pyriforma</i> (Edmondson, 1933)			
<i>Troglocarcinus</i> Verrill, 1908	Astrocoeniidae: <i>Stephanocoenia</i> Caryophylliidae: <i>Polychathu</i> Faviidae: <i>Diploria</i> , <i>Manicina</i> Meandrinidae: <i>Dichocoenia</i> Mussidae: <i>Isophyllia</i> , <i>Mussa</i> , <i>Mussimilia</i> , <i>Myce-</i> <i>tophyllia</i> , <i>Scolymia</i> Oculinidae: <i>Oculina</i> Siderastreidae: <i>Siderastrea</i>	Atlantic: Bermuda, Florida, Caribbean south to Brazil, Ascension Island, eastern Atlantic; depth <1 to 75 m	Kropp & Manning 1987
<i>Troglocarcinus corallicola</i> (Fize & Serène, 1956a)			Carricart-Ganivet et al. 2004
<i>Utinomiella</i> Kropp & Takeda, 1988	Pocilloporidae: <i>Pocillopora</i> , <i>Stylophora</i>	Pacific: Japan (Ryukyu Islands), Micronesia (Palau, Guam, Pohnpei), Hawaii Indian Ocean: Andaman Islands; depth 1 to 29 m	Kropp 1990 Wei et al. 2006
<i>Utinomiella dimorpha</i> (Henderson, 1906)			

Table 1. continued.

Genus and Species	Known Coral Hosts	General Distribution (of crab)	Primary References
<i>Xynomaia</i> Kropp, 1990	Faviidae: <i>Favia</i> , <i>Goniastrea</i> , <i>Montastrea</i> , <i>Oulophyllia</i> , <i>Platygyra</i> Merulinidae: <i>Merulina</i> Pectiniidae: <i>Pectinia</i>	Pacific: Vietnam, Sumatra, Japan (Izu Islands, Kushimoto), Micronesia (Palau, Guam); depth 1 to 15 m	Kropp 1990
<i>Xynomaia boissoni</i> (Fize & Serène, 1956a)			
<i>Xynomaia sheni</i> (Fize & Serène, 1956b)			
<i>Xynomaia verrilli</i> (Fize & Serène, 1957)			
<i>Zibrovía</i> Kropp & Manning, 1996	Phyllangiidae: <i>Phyllangia</i>	Pacific: Philippines Indian Ocean: Madagascar; depth 81 to 100 m	Kropp & Manning 1996
<i>Zibrovía galea</i> Kropp & Manning, 1996			

¹ The question mark after the genus name in *Lithoscaptus pacificus* refers to the fact that, because of the poor condition of the type of *Cryptochirus pacificus* Edmondson, Kropp (1990) placed the species in the genus *Lithoscaptus* only tentatively.

that includes all genera and species of the family, the host scleractinian coral genus from which they have been reported, and the general distribution patterns of each cryptochirid genus.

Presumably as an adaptation to their environment (their close association with corals), the cryptochirids have evolved a small, squat, and distinctive body that, although perhaps superficially similar to crabs of the family Pinnotheridae in some species, is unlike that of other crab families, even those that also live as obligate commensals of corals (e.g., trapeziids and domeciids). Based on their morphology, in the most current (and indeed in all other) classifications, the gall crabs are placed in their own family (Cryptochiridae) and superfamily (Cryptochiroidea). There is some (unpublished) information indicating that the family is probably monophyletic (Kropp 1988), but little beyond that. Even placement of the superfamily within the Eubrachyura (higher crabs) has been historically uncertain. For example, Martin & Davis (2001) placed the gall crabs within the subsection Heterotremata, whereas the most recent treatment of the Brachyura (Ng et al. 2008) places the superfamily Cryptochiroidea in the subsection Thoracotremata. It would seem, therefore, that the question of the origin and evolutionary relationships of the cryptochirid crabs is a question perfectly suited to investigation with molecular systematic techniques. We address for the first time the evolutionary relationships of gall crabs to other brachyuran families using molecular sequence data. This study must be considered preliminary in that only two populations of a single species (the widespread *Hapalocarcinus marsupialis* Stimpson, 1859) were included, but the results seem sufficiently robust to suggest affinities of the gall crabs at the superfamily and possibly family level.

2 MATERIALS AND METHODS

We sequenced a ~545-bp fragment of the 16S mitochondrial gene from Mexican and French Polynesian specimens of the cryptochirid *Hapalocarcinus marsupialis* Stimpson, 1895. The Mexican material was extracted from crabs removed from corals that had been in the collections of the Natural History Museum of Los Angeles County. The Polynesian material was collected in 2001 and was preserved in ethanol. Locality and collection details as well as GenBank numbers are included in Table 2. Muscle tissue was taken from the fifth pereopod and was extracted with a QIAGEN DNeasy Kit (Qiagen, Valencia, CA). The manufacturer's protocol was followed for extraction, and tissue was macerated in a PCR tube with a pestle and then incubated in a 55°C incubator overnight on a shaking table set to medium speed. Polymerase chain reaction (PCR, Sakai et al. 1988) was carried out with standard PCR conditions (2.5 µl of 10x PCR buffer, 1.5 µl of 50 mM MgCl₂, 4 µl of 10 mM dNTPs, 2.5 µl each of two 10 pmol primers, 0.15 Platinum *Taq* (5 units/µl), 9.6 µl double distilled water, and 1 µl template) and thermal cycling as follows: an initial denaturation at 96°C for 3 minutes followed by 40 cycles of 95°C for 1 minute, 46°C for 1 minute, and 72°C for 10 minutes. 16SrDNA was amplified in both directions with universal 16Sar and 16Sbr primers (Palumbi et al. 1991). PCR products were visualized by agarose (1.2%) gel electrophoresis with Sybr Gold (Invitrogen, Carlsbad, CA), PCR product was purified with Sephadex (Sigma Chemical, St. Louis, MO) on millipore multiscreen filter plates, and DNA was cycle sequenced with ABI Big-dye ready-reaction kit and following the standard cycle sequencing protocol with one quarter of the suggested reaction volume.

Sequences were edited and assembled in Sequencher (Gene Codes Corporation); 16S rDNA was aligned using MAFFT (Multiple Alignment Program for amino acid or nucleotide sequences, Katoh et al. 2002; Katoh et al. 2005) and manually adjusted where mismatches were made. All three LINS, EINS, and GINS alignment protocols were reviewed. Phylogenetic trees were estimated with maximum likelihood (GARLI, Genetic Algorithm for Rapid Likelihood Inference, Zwickl 2006). GARLI phylogenetic searches on aligned nucleotide datasets begin with an assumed model of nucleotide substitutions (GTR), with gamma distributed rate heterogeneity and an estimated proportion of invariable sites. The implementation of this model is exactly equivalent to that in PAUP*, making the log likelihood (lnL) scores obtained directly comparable. All model parameters were estimated, including the equilibrium base frequencies. The gamma model of rate heterogeneity

Table 2. Cryptochirids sequenced and GenBank sequences used in analyses.

Subsection	Superfamily	Family	Genus/species	GenBank No.
	Cryptochiroidea	Cryptochiridae	<i>Hapalocarcinus marsupialis</i>	EU743929
				EU743930
			Mexico, Baja California Sur, Palmas Bay, Rancho Buena Vista, <i>Pocillopora</i> with barnacles, 4.57 m. Original fixative unknown, specimen in 70% ethanol. 15 Sep. 1962. AHF, 1963-13, lot 13, cat. no. 530, JM-2005-003. Coll. Edmond Hobsen. RW05.301.1154.	
				EU743930
			Pacific, Society Islands, French Polynesia, Moorea, 6 km south of airport, site 9, ~17.533°S ~149.783°W, <i>Pocillopora</i> with barnacles, snorkel to motu, very close to outer reef, original fixative rum 50% ethanol, subsequently transferred to 95% ethanol. 25 Jul. 2001. JM-2005-004, ST01.055. Coll. Sandy Trautwein. RW05.302.1155.	
Heterotremata	Potamoidea	Gecarcinucidae	<i>Sartoriana spinigera</i>	AM234649
		Potamidae	<i>Geothelphusa pingtung</i>	AB266168
Thoracotremata	Grapsoidea	Gecarcinidae	<i>Cardisoma carnifex</i>	AM180687
			<i>Gecarcinus lateralis</i>	AJ130804
			<i>Gecarcoidea lalandii</i>	AM180684
		Glyptograpsidae	<i>Glyptograpsus impressus</i>	AJ250646
			<i>Platychoirapsus spectabilis</i>	AJ250645
		Grapsidae	<i>Geograpsus lividus</i>	AJ250651
			<i>Goniopsis cruentata</i>	AJ250652
			<i>Grapsus grapsus</i>	AJ250650
			<i>Leptograpsus variegatus</i>	AJ250654
			<i>Metopograpsus latifrons</i>	AJ784028
			<i>Metopograpsus quadridentatus</i>	DQ062732
			<i>Metopograpsus thukuhar</i>	AJ784027
			<i>Pachygrapsus crassipes</i>	AB197814
			<i>Pachygrapsus marmoratus</i>	DQ079728
			<i>Pachygrapsus minutus</i>	AB057808
			<i>Pachygrapsus transversus</i>	AJ250641
			<i>Planes minutus</i>	AJ250653
		Plagusiidae	<i>Euchirograpsus americanus</i>	AJ250648
			<i>Percnon gibbesi</i>	AJ130803
			<i>Plagusia squamosa</i>	AJ311796
		Sesarmidae	<i>Armases elegans</i>	AJ784011
			<i>Sarmatium striaticarpus</i>	AM180680
			<i>Sesarma meridies</i>	AJ621819
			<i>Sesarma windsor</i>	AJ621824
			<i>Sesarmoides longipes</i>	AJ784026
		Varunidae	<i>Austrohelice crassa</i>	AJ308416
			<i>Brachynotus atlanticus</i>	AJ278831
			<i>Cyrtograpsus affinis</i>	AJ130801
			<i>Eriocheir sinensis</i>	AJ250642
			<i>Gaetice americanus</i>	AJ250643
			<i>Helograpsus haswellianus</i>	AJ308417
			<i>Hemigrapsus oregonensis</i>	AJ250644

Table 2. continued.

Subsection	Superfamily	Family	Genus/species	GenBank No.		
Ocyphoidea	Camptandriidae		<i>Hemigrapsus sanguineus</i>	AJ493053		
			<i>Paragrapsus laevis</i>	AJ308418		
			<i>Varuna litterata</i>	AJ308419		
			<i>Baruna trigranulum</i>	AB002129		
			<i>Paracleistostoma depressum</i>	AB002128		
			Mictyridae	<i>Mictyris brevidactylus</i>	AB002133	
				Ocyphodidae	<i>Dotilla wichmanni</i>	AB002126
			<i>Ilyoplax deschampsii</i>		AB002117	
					<i>Scopimera globosa</i>	AB002125
					<i>Tmethypocoelis ceratophora</i>	AB002127
				Palicidae	<i>Crossotonotus spinipes</i>	AJ130807
					<i>Palicus caronii</i>	AM180692
Pinnotheroidea	Pinnotheridae	<i>Austinixa hardyi</i>	AF503185			
		<i>Austinixa patagoniensis</i>	AF503186			
		<i>Pinnotheres pisum</i>	AM180694			

assumes four rate categories. GARLI uses a genetic algorithm approach to simultaneously find the topology, branch lengths, and model parameters that maximize the lnL (Zwickl 2006).

The phylogeny was also estimated with Mr. Bayes 3.0b4 (Ronquist & Hulsenbeck 2003) using Bayesian inferences coupled with Markov chain Monte Carlo techniques. Four Markov–Monte Carlo chains were run for ten million generations, and a sample tree was saved every 1000 generations. Trees chosen from the first one million generations were discarded as “burn in.” Trees that were chosen once likelihood scores converged on a stable value were used to construct a 50% majority rule consensus tree in PAUP*.

A ~1860-bp double-stranded fragment of 18SrDNA was also sequenced but not used due to a lack of sequence variation (GenBank numbers EU743931 and EU743932). Taxon selection for the analyses was repeatedly refined, as it was determined that Cryptochiridae are members of Thoracotremata and the Grapsoidea and are nested within the Grapsidae. This realization changed our approach from focusing on 18S rDNA to the more appropriate 16S rDNA for this analysis. Taxa selected for the 16S dataset included broad, but not exhaustive, sampling of Varunidae, Grapsidae, Plagusidae, Sesarnidae, Camptandriidae, Gecarcinidae, Pinnotheridae, and Mictyridae, with the goal of associating the Cryptochiridae with its closest relatives.

3 RESULTS

Analyses of our cryptochirids from Mexico and Polynesia revealed that despite their geographic separation, both samples were the same species, the widespread and relatively common *Hapalocarcinus marsupialis* Stimpson, 1859. In all of our analyses, the cryptochirids are nested within a group of crabs considered by most workers to constitute the Thoracotremata. More specifically, the genus *Hapalocarcinus* falls within a clade that includes the familiar grapsid genera *Grapsus*, *Geograpsus*, *Goniopsis*, *Leptograpsus*, *Planes*, and *Pachygrapsus* (Fig. 1). Branch lengths for the two *Hapalocarcinus* sequences are long, as is the branch length of the *Mictyris* sequence (not shown). Interestingly, however, *Hapalocarcinus* was not close to some of the grapsoids that are common reef inhabitants, such as the genera *Percnon* and *Plagusia*, both of which were at one time considered members of the family Plagusidae (but see below). Beyond our observations on the gall crabs (based on this single species), our results also indicate that the genus *Pachygrapsus* is not monophyletic, with *P. marmoratus* not clustering with the other four *Pachygrapsus* species.

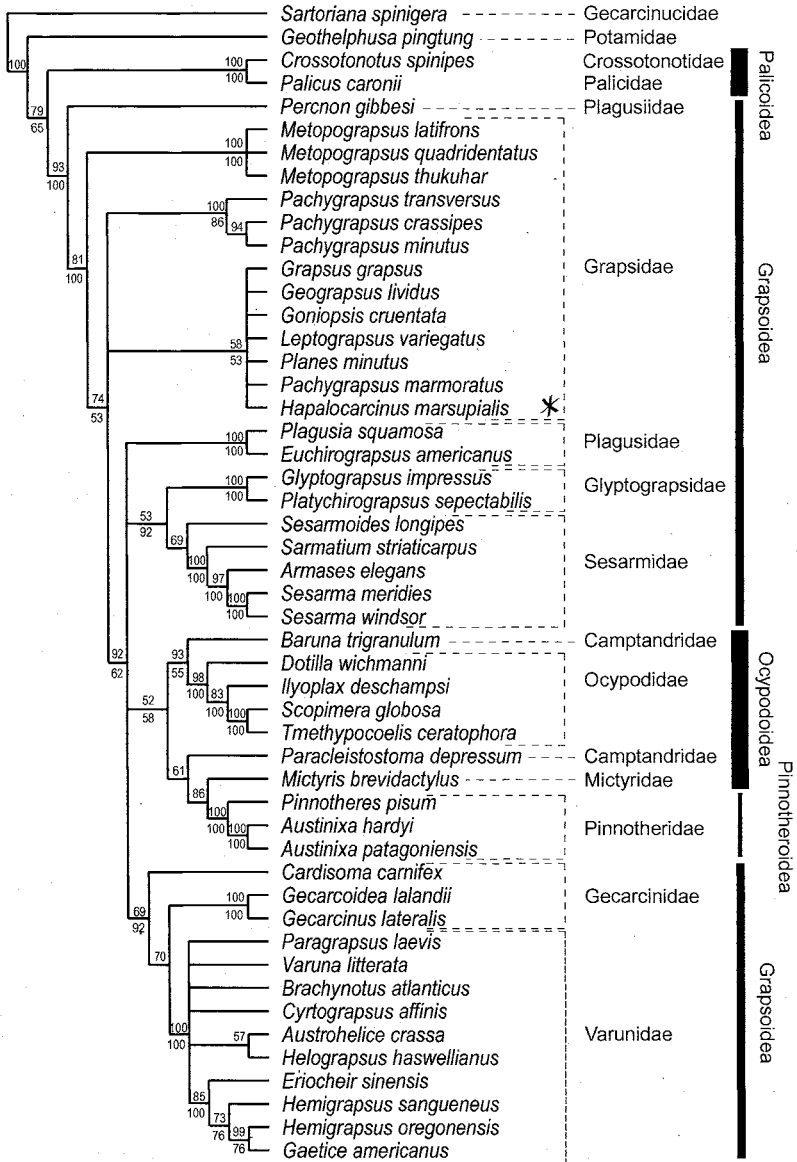


Figure 1. Phylogenetic placement of the Cryptochiridae, represented by the genus *Hapalocarcinus* (*), and relationships of Ocypodoidea, Grapsoidea, Pinnotheroidea, and Palicoidea based on 16S mtDNA sequences of 51 taxa, 589 characters, nucleotide frequencies: f(A) = 0.24387, f(C) = 0.24433, f(G) = 0.27220, f(T) = 0.23960. This tree is rooted in Gecarcinidae and Potamidae. Topology derived from Bayesian inference 50% majority rule consensus of 18,000 trees. Significance values are posterior probabilities >50% above the branches. GARLI maximum likelihood ln score = -8935.92, 50% majority rule consensus of 74 trees; bootstrap values are below the branches.

Maximum likelihood and Bayesian analyses converged on the same topology. All of our analyses recognize *Glyptograpsus* and *Platychirograpsus* as sister taxa, confirming their placement in the family Glyptograpsidae. The species of *Pinnotheres* and *Austinixa* selected for this analysis constitute a monophyletic clade (the Pinnotheridae). The Varunidae (*Austrohelice*, *Brachynotus*,

Cyrtograpsus, *Eriocheir*, *Gaetice*, *Helograpsus*, *Hemigrapsus*, *Paragrapsus*, and *Varuna*) is a well-supported monophyletic clade. Gecarcinidae are basal to the Varunidae (posterior probability 69% and bootstrap support 92%). As alluded to above, the plagusiid genera *Plagusia* and *Euchirograpsus* are sister taxa, but they are not at all closely related to the genus *Percnon*, previously included in the Plagusiidae.

At the superfamily level, Pinnotheroidea appears monophyletic, although only three taxa were used in our analysis. The Palicoidea appears as monophyletic and basal to the “grapsoids” in our phylogeny. In our analysis, the superfamilies Ocypodoidea and Grapsoidea are not monophyletic clades.

4 DISCUSSION

As noted earlier, in all of our analyses, which must be considered preliminary because of the single species used to represent the gall crabs, the cryptochirids are nested within a group of crabs considered by most workers to constitute the Thoracotremata. This group is defined primarily by having the location of the opening of the vas deferens through the sternum rather than through the coxa of the fifth pereopod (Ng et al. 2008: 8). This placement agrees with the most recent compilation and classification of crabs by Ng et al. 2008 and not with the classification suggested by Martin & Davis (2001), in which the cryptochirids were treated as members of the more diverse Heterotremata. The Ng et al. (2008) classification treats the Thoracotremata as being composed of 17 extant families distributed among four superfamilies: Cryptochiroidea, Grapsoidea, Ocypodoidea, and Pinnotheroidea.

Within the Thoracotremata, our best tree places the gall crab genus *Haplocarcinus* within a clade that includes the familiar grapsid genera *Grapsus*, *Geograpsus*, *Goniopsis*, *Leptograpsus*, *Planes*, and *Pachygrapsus*. Since only a single species was sampled in the family, the long branch length of *Haplocarcinus* precludes more accurate placement within the grapsids in this analysis. The association of *Haplocarcinus* with grapsid genera is a somewhat surprising result, in part because there are other groups of crabs that are closely associated with reefs (e.g., trapeziids, domeciids, and some other coral-associated taxa). Also surprising to us was that, even among grapsoids, there are genera more typically associated with reef-dwelling than those with which *Haplocarcinus* clusters, such as *Percnon* and *Plagusia*; these were not close to the gall crabs in our results. The transition from a coral-obligate commensal group of crabs (such as the trapeziids, tetraliids, or domeciids) to a more heavily coral-dependent group such as the gall crabs would have been, in some ways, easier to understand. However, no such coral-obligates are seen among the crabs that appear closest to *Haplocarcinus* in our analysis. We should also point out that adaptation to a coral-associated lifestyle does not always result in similar modifications, even among decapods (e.g., consider the morphological differences between trapeziids and domeciids such as *Maldivia*, or between the shrimp genera *Paratypton* and *Alpheus*) despite similar lifestyles and diets.

Some traditional groupings, such as the families Varunidae, Pinnotheridae, Ocypodidae, Sesarmidae, and Glyptograpsidae, are supported in this analysis. However, other traditionally recognized families, such as the Camptandriidae and Plagusiidae, are not supported (see also Schubart et al. 2002; Schubart et al. 2006). Although a case could be made for recognition of the superfamily Pinnotheroidea, and possibly the Ocypodoidea (with the exception of the genera *Paracleistostoma* and *Mictyris*), there is no support for the superfamilies Cryptochiroidea, Grapsoidea, and Ocypodoidea as previously defined (Fig. 1). This perhaps is not surprising in light of the rather weak and likely convergent morphological characters that have been used to define these superfamilies in the past (such as the “rectangular” carapace shape of the grapsoids and the long eyestalks of many ocypodoids).

The pinnotherids, all of which are highly modified (most having extremely short and wide bodies) for a commensal existence, appear to be monophyletic and are not closely related to cryptochirids despite an apparently superficial resemblance (see Introduction), although this result is

based on only three representatives of that family. The former family Palicidae (*Crossotonotus* + *Palicus*) (now treated as two families, Crossotonotidae and Palicidae, within a superfamily Palicoidea; Ng et al. 2008) appears basal to the other (non-outgroup) crabs in our study. Palicids are morphologically very unusual in that they have greatly reduced fifth pereopods (see Castro 2000).

Our results are in general agreement with the findings of Schubart et al. (2002, 2006) in their studies of the Glyptograpsidae and of the relationships within the Grapsoidea, respectively. As in the conclusion of Schubart et al. (2006), our results cast doubt on the usefulness of the superfamily categories Grapsoidea and Ocyphodoidea, and confirm that *Percnon* is not allied to *Plagusia* and *Euchirograpsus*, such that the family Plagusiidae cannot be recognized as monophyletic.

For the gall crabs, the superfamily status of the Cryptochiroidea is now difficult to justify, as, based on our admittedly small dataset, the gall crabs appear to be highly modified grapsids. For practical reasons, and until more cryptochirid sequences from a broader family sampling are included in future analyses, we suggest maintaining the family status of the Cryptochiridae but treating it as one of many separate “grapsoid” families. We recommend dropping the superfamily category (Cryptochiroidea), while at the same time recognizing that the Grapsoidea, as previously defined, is itself an artificial assemblage. The rather wide geographical range of the gall crabs, summarized in Table 1, and the fact that, despite the geographical distance between the populations sampled in this study (Mexico and French Polynesia), our sequences came from a single species, also are reasons to suspend making any higher-level classificatory changes, as it is possible that convergence to a coral-dwelling habitat has occurred more than once.

ACKNOWLEDGEMENTS

This work was supported in part by grant number DEB 0531616 from the National Science Foundation’s “Assembling the Tree of Life” program to J. W. Martin, in conjunction with collaborative awards to Keith Crandall and Nikki Hannegan (Brigham Young University), Darryl Felder (University of Louisiana Lafayette), and Rodney Feldmann and Carrie Schweitzer (Kent State University). The symposium during which these results were first presented was funded by NSF grant DEB 0721146, with additional support from the American Microscopical Society, the Crustacean Society, the Society of Systematic Biologists, the Society of Integrative and Comparative Biology (SICB), and the SICB Divisions of Invertebrate Zoology and Systematics & Evolutionary Biology. We sincerely thank Roy Kropp and Gustav Paulay for providing information and literature on gall crab biology and for helpful reviews of the manuscript. The participation of J. Martin was also made possible by funding from the U.S. National Oceanographic and Atmospheric Administration NOAA for systematic work on crabs of the Hawaiian Islands. Additional support (for R. Wetzer) was provided by NSF grant DEB-0129317. We especially thank Keith Crandall and the members of his laboratory, especially Rebecca Scholl, Katharina Dittmar, Marcos Pérez-Losada, and Megan Porter, for their hospitality, assistance, and mentoring at the bench as well for their help with the analyses during R. Wetzer’s working visits to Brigham Young University in 2006–2007.

REFERENCES

- Abelson, A., Galil, B.S. & Loya, Y. 1991. Skeletal modifications in stony corals caused by in-dwelling crabs: hydrodynamic advantages for crab feeding. *Symbiosis* 10: 233–248.
- Carricart-Ganivet, J.P., Carrera-Parra, L.F., Quan-Young, L.I. & Garia-Madrigal, M.S. 2004. Ecological note on *Troglocarcinus corallicola* (Brachyura: Cryptochiridae) living in symbiosis with *Manicina areolata* (Cnidaria: Scleractinia) in the Mexican Caribbean. *Coral Reefs* 23: 215–217.

- Castro, P. 2000. Crustacea Decapoda: A revision of the Indo-West Pacific species of palicid crabs (Brachyura Palicidae). In: Crosnier, A. (ed.), *Résultats des Campagnes Musorstom* 21: 437–610.
- Edmondson, C.H. 1925. Marine zoology of tropical central Pacific. Crustacea. *Bull. Bernice Bishop Mus.* 27: 3–62.
- Edmondson, C.H. 1933. *Cryptochirus* of the central Pacific. *Bernice Bishop Mus. Occas. Papers* 10: 1–23.
- Fize, A. & Serène, R. 1957. Les Hapalocarcinidés du Viêt-Nam. *Mém. l'Institut Océanograph. Nhatrang* 10: 1–202.
- Fize, A. & Serène, R. 1956a. Note préliminaire sur huit espèces nouvelles, dont une d'un genre nouveau, d'Hapalocarcinidés. *Bull. Soc. Zool. France* 80: 375–378.
- Fize, A. & Serène, R. 1956b. Note préliminaire sur quatre espèces nouvelles d'Hapalocarcinidés avec quelques remarques au sujet du *Cryptochirus rugosus* Edmondson. *Bull. Soc. Zool. France* 80: 379–382.
- Heller, C. 1861. Synopsis der im rothen Meere vorkommenden Crustaceen. *Verhandl. Zoolog.-Botan. Gesellsch. Wien* 11: 3–32.
- Henderson, J.R. 1906. On a new species of coral-infesting crab taken by the R.I.M.S. 'Investigator' at the Andaman Islands. *Ann. Mag. Nat. Hist. Ser. 7* 18: 211–219.
- Hiro, F. 1938. A new coral-inhabiting crab, *Pseudocryptochirus viridis* gen. et. sp. nov. (Hapalocarcinidae, Brachyura). *Zool. Magazine, Tokyo* 50: 149–151.
- Katoh, K., Kuma, K., Toh, H. & Miyata, T. 2005. MAFFT version 5: improvement in accuracy of multiple sequence alignment. *Nucleic Acids Res.* 33: 511–518.
- Katoh, K., Misawa, K., Kuma, K., & Miyata, T. 2002. MAFFT: a novel method for rapid multiple sequence alignment based on fast Fourier transform. *Nucleic Acids Res.* 30: 3059–3066.
- Kropp, R.K. 1986. Feeding biology and mouthpart morphology of three species of coral gall crabs (Decapoda: Cryptochiridae). *J. Crust. Biology* 6: 377–384.
- Kropp, R.K. 1988. Biology and systematics of coral gall crabs (Crustacea: Cryptochiridae). Ph.D. dissertation, University of Maryland, College Park. 354 pp.
- Kropp, R.K. 1989. A revision of the Pacific species of gall crabs, genus *Opecarcinus* (Crustacea: Cryptochiridae). *Bull. Mar. Sci.* 45: 98–129.
- Kropp, R.K. 1990. Revision of the genera of gall crabs (Crustacea: Cryptochiridae) occurring in the Pacific Ocean. *Pacific Sci.* 44: 417–448.
- Kropp, R.K. 1994. The gall crabs (Crustacea: Decapoda: Brachyura: Cryptochiridae) of the Rumphius Expeditions revisited, with descriptions of three new species. *Raffles Bull. Zool.* 42: 521–538.
- Kropp, R.K. 1995. *Lithoscaptus pardalotus*, a new species of coral-dwelling gall crab (Crustacea: Brachyura: Cryptochiridae) from Belau. *Proc. Biol. Soc. Washington* 108: 637–642.
- Kropp, R.K. & Manning, R.B. 1987. The Atlantic gall crabs, family Cryptochiridae (Crustacea: Decapoda: Brachyura). *Smithson. Contrib. Zool.* 462: 1–21.
- Kropp, R.K. & Manning, R.B. 1996. Crustacea Decapoda: two new genera and species of deep water gall crabs from the Indo-west Pacific (Cryptochiridae). In: Crosnier, A. (ed.), *Résultats des Campagnes MUSORSTOM*, vol. 15. *Mém. Mus. Nat. His. Natur., Paris* 168: 531–539.
- Kropp, R.K. & Takeda, M. 1988. *Utinomiella*, a replacement name for *Utinomia* Takeda et Tamura, 1981 (Crustacea, Decapoda), non Tomlinson, 1963 (Crustacea, Acrothoracica). *Bull. Biogeog. Soc. Japan* 43: 29.
- Manning, R.B. 1991. Crustacea Decapoda: *Cecidocarcinus zibrowii*, a new deep water gall crab (Cryptochiridae) from new Caledonia. In: Crosnier, A. (ed.), *Résultats des Campagnes MUSORSTOM*, vol. 9. *Mém. Mus. Nat. His. Natur., Paris* 152: 515–520.
- Martin, J.W. & Davis, G.E. 2001. An updated classification of the Recent Crustacea. *Natural History Museum of Los Angeles County, Science Series* 39: 1–124.

- Milne Edwards, A. 1862. (Annexe F) Faune carcinologique de l'île de las Réunion. In: Maillard, L. (ed.), *Notes sur l'île de la Réunion*. Paris. 1–16.
- Monod, T. 1956. Hippidea et Brachyura ouest-africains. *Mém. Institut Franç. Afrique Noire* 45: 1–674.
- Ng, P.K.L., Guinot, D. & Davie, P.J.F. 2008. Systema Brachyurorum: Part I. An annotated checklist of extant brachyuran crabs of the world. *Raffles Bull. Zool.* 17: 1–286.
- Palumbi, S.R., Martin, A., Romano, S., McMillan, W.O., Stice, L. & Grabowski, G. 1991. *The Simple Fool's Guide to PCR, version 2*. Department of Zoology and Kewalo Marine Laboratory, University of Hawaii, Honolulu, Hawaii. 43 pp.
- Paul'son, O.M. 1875. Izsledovaniya rakoobraznykh krasnago morya s zametkami otnositel'no rakoobraznykh drugikh morei. Chast' 1. Podophthalmata i Edriophthalmata (Cumacea). S.V. Kul'zhenko, Kiev. 164 pp.
- Paulay, G., Kropp, R., Ng, P.K.L. & Eldredge, L.G. 2003. The crustaceans and pycnogonids of the Mariana Islands. *Micronesica* 35–36: 456–513.
- Ronquist, F. & Huelsenbeck, J.P. 2003. MrBayes 3: Bayesian phylogenetic inference under mixed models. *Bioinformatics* 19: 1572–1574.
- Sakai, R., Gelfand, D.J., Srofeff, S., Scharf, S.J., Higuchi, R., Horn, G.T., Mullis, K.B. & Erlich, H.A. 1988. Primer-directed enzymatic amplification of DNA with a thermostable DNA polymerase. *Science* 239: 487–491.
- Schubart, C.D., Cannicci, S., Vannini, M. & Fratini, S. 2006. Molecular phylogeny of grapsoid crabs (Decapoda, Brachyura) and allies based on two mitochondrial genes and a proposal for refraining from current superfamily classification. *J. Zool. System. Evolut. Res.* 44: 193–199.
- Schubart, C.D., Cuesta, J.A. & Felder, D.L. 2002. Glyptograpsidae, a new brachyuran family from Central America: larval and adult morphology, and a molecular phylogeny of the Grapsoidea. *J. Crust. Biol.* 22: 28–44.
- Serène, R. 1966. Sur deux espèces nouvelles de Brachyours (Crustacés Décapodes) et sur une troisième peu connue, récoltées dans la région malaise. *Bull. Mus. Nat. Hist. Nat., Paris* 38: 817–827.
- Shaw, K. & Hopkins, T.S. 1977. The distribution of the family Hapalocarcinidae (Decapoda, Brachyura) on the Florida Middle Ground with a description of *Pseudocryptochirus hypostegus* new species. *Proc. 3rd Internat. Coral Reef Symp., Miami* 1: 177–183.
- Shen, C.J. 1936. Notes on the family Hapalocarcinidae (coral-infesting crabs) with description of two new species. *Hong Kong Nat. Suppl.* 5: 21–26.
- Stimpson, W. 1859. [Communication, *Hapalocarcinus marsupialis*]. *Proc. Boston Soc. Nat. Hist.* 6: 412–413.
- Takeda, M. & Tamura, Y. 1980a. Coral-inhabiting crabs of the family Hapalocarcinidae from Japan. *Bull. Nat. Sci. Mus., Tokyo, Series A (Zoology)* 3: 147–151.
- Takeda, M. & Tamura, Y. 1980b. Coral-inhabiting crabs of the family Hapalocarcinidae from Japan, III. New genus *Fizesereneia*. *Bull. Nat. Sci. Mus., Tokyo, Series A (Zoology)* 6: 137–146.
- Takeda, M., & Tamura, Y. 1981a. Coral-inhabiting crabs of the family Hapalocarcinidae from Japan. VIII. Genus *Pseudocryptochirus* and two new genera. *Bull. Biogeog. Soc. Japan* 36: 13–27.
- Takeda, M. & Tamura, Y. 1981b. Coral-inhabiting crabs of the family Haplocarcinidae from Japan. VII. Genus *Favicola*. *Res. on Crustacea, Carcin. Soc. Japan* 11: 41–50.
- Takeda, M. & Tamura, Y. 1983. Coral-inhabiting crabs of the family Hapalocarcinidae from Japan. IX. A small collection made at Kushimoto and Koza, Kii Peninsula. *Bull. Nat. Sci. Mus., Tokyo, Ser. A (Zoology)* 9: 1–12.

- Verrill, A.E. 1908. Decapod Crustacea of Bermuda; 1. Brachyura and Anomura. Their distribution, variations, and habits. *Trans. Connecticut Acad. Arts Sci.* 13: 299–473.
- Wei, T.-P., Hwang, J.-S., Tsai, M.-L. & Fang, L.-S. 2006. New records of gall crabs (Decapoda, Cryptochiridae) from Orchid Island, Tawain, Northwestern Pacific. *Crustaceana* 78: 1063–1077.
- Zwickl, D.J. 2006. Genetic algorithm approaches for the phylogenetic analysis of large biological sequence datasets under the maximum likelihood criterion. Ph.D. dissertation, The University of Texas at Austin, Austin, TX.